The dual role of phenolic compounds in oxidative changes in fruit products

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<u>Abstract</u>

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This study was conducted to reconcile the controversial scientific evidence indicating the dual role of phenolic compounds in oxidative changes in fruit products. The hypothesis has been substantiated, according to which the functioning of flavonoids as redox-systems, is strictly connected with the ability of fruit phenolic compounds to inhibit the free-radical processes of the oxidation of substrates; this is due to the semiquinone presence in the equilibrium system. Terminology and appropriate methods of studying, in respect of the antioxidant effect of natural, as well as added compounds in fruit products, were analysed. The effect of redox-properties of the antioxidant agent, on studying the inhibition of the enzymatic oxidation of phenolic compounds with this agent, has been analysed. It was shown that this field of study is vital for improving the concepts of browning inhibitors in foodstuffs, as well as for improving the theory of interactions between components in canned foodstuffs.

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Introduction

Terms in respect of fruit phenolic compounds

Depending on the type of carbon skeletons, fruit phenolic substances were divided into three groups: flavonoids with $C_6-C_3-C_6$ structure; cinnamic acid derivatives with $C_6 - C_3$ structure; phenolcarbonic acids with C_6-C_1 structure (Zaprometov, 1964). The term "fruit polyphenols" is used to define phenolic compounds in their monomeric forms such as cyanidin; pelargonidin, etc. and their glycosides (Sioud and Luh, 1966; Scoricova, 1973; Tsao, 2010; Watson et al., 2014). Contemporary authors also call them polyphenolic substances (Kratchanova, 2010; Georgieva and Mihaylova, 2015), phenolic compounds (Martinez and Whitaker, 1995; Zheng and Wang, 2001; Balasundrama et al., 2006; Mertz et al., 2007; Wojdylo et al., 2007; Huand et al., 2010; Cheynie, 2012), and polyphenolic compounds (Handique and Baruah, 2002; Ignat et al., 2011; Sasikumar et al., 2015; Sripakdee et al., 2015). The chemical structure of flavonoids allows them to be classified as diphenols. The term 'polyphenol' is historically used and is well matched with 'polyphenol oxidase (PPO)', which is used for the enzyme catalysing oxidation process in plants, and fruit pulps (Yoruk and Marshall, 2003; Kolodziejczyk et al., 2010). Therefore, the terms 'phenolic compound', and 'polyphenol' are both acceptable for describing the monomeric forms of flavonoids, cinnamonic acid derivaties, and phenolcarbonic acids. Today, the term 'polyphenol' not only includes phenolic acids, and flavonoids, but also stilbenes, lignans, and polymeric lignans (Gharras, 2009; Bahadoran *et al.*, 2013; Keerthi *et al.*, 2014). The polymeric forms of fruit phenolic compounds are often referred to as tannins (Martinez and Whitaker, 1995; Caballero, 2013).

Comparative characteristics and terms in respect of the antioxidant effect of fruit phenolic compounds

The antioxidant effect of fruit phenolic substances has been extensively studied (Van Acker *et al.*, 1996; Calado *et al.*, 2015; Georgieva and Mihaylova, 2015; Korotkova *et al.*, 2015). The common definition 'antioxidant' is used for substances, which prevent or inhibit the oxidation process (Choe and Min, 2009; Flora, 2009). The antioxidant effect has been defined as an ability to protect against free radicals (Georgieva and Mihaylova, 2015). The modern quantitative methods of antioxidant activity evaluation are based on free radical absorbance capacity (Fedina *et al.*, 2010; Korotkova *et al.*, 2015).

The classical definition of antioxidants, according to which antioxidants must be readily oxidized (Housecroft and Constable, 2006) will be used in this article. Such substances include hydroquinone (Housecroft and Constable, 2006). The substances with low redox potential are classical antioxidants (Vetter, 1967). The classical antioxidant used in the fruit canning industry is ascorbic acid (Ramaswamy and Marcotte, 2005; Flaumenboum *et al.*, 2006; Adria, 2009). The mechanism of antioxidant action includes transforming the antioxidant into its oxidized form while other substances convert from its oxidized form into reduced form. For such an ability of antioxidants there is another definition referred to as the reducing power (Georgieva and Mihaylova, 2015). There is data of measuring this parameter in comparison with vitamin C (Georgieva and Mihaylova, 2015).

As can be seen from the above, there is not a unified approach to the characterisation of the antioxidant properties of phenolic substances of fruit products. However, there is no doubt about the antioxidant properties of cathechins as redox systems, and as an inhibitor of free radical oxidation of substrates (Zaprometov, 1964; Akannia *et al.*, 2014; Preedy, 2014).

Negative role of fruit phenolic compounds and methods for reducing oxidative browning

It has been shown that among the main negative changes, which take place in raw fruit materials during processing, there are oxidative polyphenol conversions, and polymerisation of products of oxidative reaction (Marh et al., 1985; Sharma et al., 2002; Boekel et al., 2010). Indeed, the damage to cell structures during processing leads to a stopping of the Palladin scheme. The interaction between: phenolic compounds, enzymes, and oxygen, during grinding, heating, or freezing of raw materials leads to the forming of brown pigments. It has been shown that dark-coloured compounds can be formed due to the interactions between guinones with proteins (Scoricova, 1973). It has been demonstrated that the monomeric forms of plant phenolic substances, such as catechins, and anthocyanins, affect negative colour changes in fruit products during processing and storage (Manach et al., 2004; Brownmiller, 2008). The browning of canned fruit products, because of non-enzymatic oxidation of phenolic compounds, has been shown to be an essential part in quality deterioration during storage (Sharma et al., 2002; Bharate, 2014).

Therefore, the oxidative browning of food products is an inevitable process caused by the nature of these foodstuffs. The importance of organoleptic properties of foodstuffs causes the fact that scientists today are still investigating new methods and new substances to reduce oxidative processes in fruit products (Dyidogan and Bayindirli, 2004; Gacche et al., 2004; Suh and Park, 2011; Wu, 2014).

Among these methods are the removal of

phenolic compounds by β -cyclodextrins (Osuga *et al.*, 1994), or by ultrafiltration (Alper and Acar, 2004), as well as eliminating phenolic compound activity by methylation (Bezusov *et al.*, 2002). However, the changes in chemical structure could affect the polyphenols' capacity to protect against free-radical oxidation of substrates.

Among methods, which could reduce oxidative processes, the most effective are inactivating the plant enzyme systems, and adding antioxidants (Thutnham, 1992; Pizzocaro, 1993; Leopoldini et al., 2011). It was reported on the heat inactivation of PPO (1, 2-benzenediol: oxygen oxidoreductase) by applying temperatures of >50°C (Martinez and Whitaker, 1995). There are recommendations of using the antioxidant agents, such as glutathione, to prevent browning in different products, including apple juice (Gacche et al., 2004), grape juice (Wu, 2014); meat products (Wu, 2013), and white wine (El Hosry et al., 2009). The authors explanations are different in respect of the positive effect of glutathione on browning inhibition, and include the antioxidant properties (Kishkovsky and Skurihin, 1988; El Hosry et al., 2009) of glutathione as well as the inhibition of the enzyme activity by glutathione (Gacche et al., 2004). It was reported on the capacity of glutathione to prevent both enzymatic and non-enzymatic browning (Wu, 2013, 2014). The controversies in explanations of the browning inhibition data with glutathione show the difficulties of investigation of PPO activity inhibition in the case of using antioxidants as enzyme inhibitors.

The importance of the investigation of the real process can be proved by the fact that the incorrect conclusion in respect of substance to be the PPO inhibitor can cause a high level of browning of canned foodstuffs processed at low heat level.

Positive role of fruit phenolic compounds

The positive effect of plant phenolic compounds was mentioned by many authors (Block and Patterson, 1992; Van Acker *et al.*, 1996; Ren *et al.*, 2003; Siriwoharn *et al.*, 2004; Ivanova *et al.*, 2005; Katalinic *et al.*, 2006; Celiktar *et al.*, 2007; Heinonen, 2007; Wojdylo *et al.*, 2007; Céspedes, 2008; Atawodi *et al.*, 2009; Dai and Mumper, 2010; Girones-Vilaplana, 2012).

The wide spectrum of the positive role of phenolic compounds could be recognized according to the following facts: first, diatomic phenols can inhibit the processes of liquid-phase oxidation of carbohydrates (Nikolaevskiy, 1978); second, there are facts about positive effect of phenols on the inhibition of the processes of enzymatic oxidation of substrates (Krylov *et al.*, 1993; Araji *et al.*, 2014); third, it has been proved the antioxidant capacity of fruit polyphenolic substances (Pastrana-Bonilla *et al.*, 2003; Du and Ma, 2009; Ramful et al. 2011; Murillo, 2012; Sasikumar *et al.*, 2015); fourth, our previous data (Bocharova, 2008) showed the positive effect of phenolic compounds of citrus fruits on vitamin C stability, and in this way conserving the biological value of products. Previous data (Bocharova *et al.*, 2016) showed that reducing ability of phenolic compounds of fruit juices redox systems affected different types of spontaneous redox reactions, which is vital for fruit foodstuffs containing benzoates in respect of the products of benzoic acid reduction.

Such a scientific controversy, in respect of the positive and negative roles of phenolic compounds in fruit products quality formation, influences the necessity of detailed analysis of this subject. The methods of preventing oxidative browning should not cause deterioration of the positive properties of fruit phenolic compounds. This substantiates the importance of the above mentioned objective.

Research task formulation

As can be seen from the above, the contradiction between the positive and negative effects of phenolic compounds on the quality of fruit products necessitates the forming of a hypothesis, which could reconcile the controversial scientific evidence. The conflicting new data necessitates forming a hypothesis on studying the browning prevention in fruit products, with respect to using an antioxidant agent as an oxidative enzyme inhibitor. This field of study is especially important for providing the competitiveness of fruit foodstuffs, as well as for establishing a scientific base for fruit products manufacturing.

To achieve the objectives of the study, it is necessary to perform the following tasks:

(1) to analyse the role of phenolic compounds as participants of oxidative changes in fruit products;

(2) to analyse the possible connection between the functioning of flavonoids as redox-systems and the ability of plant phenolic compounds to inhibit the free-radical processes of oxidation of substrates;

(3) to analyse the possibility of glutathione to be the inhibitor of oxidative enzymes;

(4) to formulate the recommendation for hypothesis-testing research.

Discussion

Fruit phenolic compounds as participants of oxidative changes in food products

Principal possibility for phenolic compounds to



Figure 1. Transformation of catechins into quinone forms

have redox properties

Fruit phenolic compounds can have conjugated bonds with oxygen atoms. It is possible to establish the reversible redox potentials in organic systems, in which there are molecules with conjugated bonds with oxygen atoms or amino groups (Vetter, 1967). It was shown that section of substances can be converted into during the reduction process.

Similar redox-reactions have been determined for phenolic substances of fruits (Zaprometov, 1964). The first product of oxidation of catechin is quinone (Zaprometov, 1964; Martinez and Whitaker, 1995; Araji *et al.*, 2014). Therefore, oxidation can lead to quinones formation in fruit products (Figure 1).

The phenolic compounds with low standard redoxpotential as possible reducing agents for some components of food system

The oxidation-reduction reactions must be in accordance with the thermodynamic abilities of the processes (Housecroft and Constable, 2006). The spontaneous processes in a system with different redox couples, are the oxidation of reduced forms of substance of redox-couple with more negative standard reduction potential (Eh_0), while oxidized forms of substance of redox-couple with more positive Eh_0 are reduced (Housecroft and Constable, 2006).

It was demonstrated in our previous work (Bocharova, 2008), that the polarisation curves of citrus juice redox-systems are situated in more negative region of potential in comparison with that of the ascorbic acid/dehydroascorbic acid couple. This explains the long stability of vitamin C in orange juice (Tressler and Jocelyn, 1957; Hui, 2006) and proves the idea about the possibility of diatomic phenols to be the inhibitors in the processes of liquid-phase oxidation of carbohydrates (Nikolaevskiy *et al.*, 1978).

To prove ours hypothesis it is necessary to study the standard redox-potentials of different flavonoids, such as hesperidin, naringenin, eriodictiol etc.

Phenolic compounds with high standard redoxpotential as substances that need antioxidants

On the contrary, the high Eh_0 of catechins

Table 1. Normal redox potentials in aqueous solutions (Kishkovsky and Skurihin 1988)

The electrode	Eh₀ (mV) at pH	
	3	7
Glutathione reduced/ glutathione oxidized	_	40
S0 ₂ / S0 ₄ ²⁻	200	_
Ascorbic acid/dehydroascorbic acid	210	_
Anthocyans reduced/ anthocyans oxidized	270	_
Tartaric acid/ dihydroxyfumaric acid	220	-
Dihydroxyfumaric acid/ diketosuccinic acid	_	300
Catechins reduced/ Catechins oxidized	430	_

(Table 1) shows the necessity of using antioxidants to prevent their conversion into an oxidized form. Such a preventer could be ascorbic acid (Table 1) or other redox-couples with more negative Eh0. The reducing of oxidized forms of catechins while ascorbic acid is converting into dehydroascorbic acid is thermodynamically favourable reaction (Table 1) and can explain the antioxidant effect of ascorbic acid on catechins. Indeed, we can see from Table 1 (Kishkovsky and Skurihin, 1988), that Eh_o of ascorbic acid/dehydroascorbic acid couple is less positive than that for catechins reduced/catechins oxidized couple. Therefore, the process of converting quinones into their reduced forms by using ascorbic acid is spontaneous, and this effect results in the inhibition of browning processes in fruit products (Scoriciva, 1973).

As can be seen from above, when food system enzymes are inactivated, the product quality will be mainly affected by the direction of redox-reactions. The role of phenolic compounds in fruit products, with respect to its redox-properties, is caused by redox-potentials. Actually, certain fruit phenolic compounds can provide vitamin C stability, and have an antioxidant effect, but some need antioxidants added to prevent their oxidation conversions into quinones, the polymerisation process, and deterioration of the colour of fruit products.

The inhibition of non-enzymatic oxidative processes with antioxidant agents in fruit products

As can be seen from the above, the non-enzymatic browning, which is connected with phenolic compound oxidation, includes:

1) oxidation of phenolic compounds to quinones (Martinez and John, 1995);

2) further transformation of quinones, including the polymerization process.

The presence of a reducer, such as ascorbic acid,

or glutathione, in food systems due to their redoxproperties leads to the opposite process of converting quinones into reduced forms. The authors reported about non-enzymatic browning inhibition with glutathione (El Hosry *et al.*, 2009; Wu, 2014). To prove the mechanism of the inhibition of browning in fruit products with glutathione, the standard redoxpotential of glutathione should be lower than the standard redox potential of fruit phenolic compounds. As can be seen from Table 1, the value of the standard redox-potential of glutathione is significantly lower than that for catechins. Therefore, the antioxidant effect of glutathione is not in doubt.

Analysing the effect of an antioxidant agent on redoxreactions in fruit systems containing PPO

The rate of oxidative processes can be enhanced with catalysts (Nelson and Cox, 2008). Such a catalyst for oxidative changes of phenolic compounds in fruit products is PPO. The class name of this enzyme is oxidoreductases. Enzymes of this class catalyse the transfer of electrons (Nelson and Cox, 2008), and provide for the conversion of phenolic compounds into quinones (Araji *et al.*, 2014). PPOs are found in different higher plants, including fruits (Siddiq and Cash, 2000; Jang and Moon, 2011; Kasikci and Bagdatlioglu, 2016), vegetables (Vamos, 1981; Hunt *et al.*, 1993; Vitti *et al.*, 2011; Hui and Evranuz, 2015), tea (Martinez and Whitaker, 1995).

The effect of substrate concentration on the velocity of an enzyme-catalysed reaction is a fundamental truth in classical biochemistry (Nelson and Cox, 2008). Therefore, the velocity of enzymecatalysed reactions increase when antioxidant agents, such as ascorbic acid or glutathione increase the amounts of substrate for PPO, because of reducing the quinones forms of diphenols into its reduced forms (Hutchings, 1994). The velocity of reduction oxidized forms of polyphenols, with an antioxidant agent, can be so significant that this reaction prevails under the reaction of enzymatic oxidation. Such an effect can be illustrated by the popular method of adding ascorbic acid before juicing. In this process, the predominance of the non-enzymatic reduction of quinones over the enzymatic oxidation of diphenols was demonstrated. The velocities of both processes are reflected in the overall direction of the reaction. Such a predominance is possible due to the saturation effect of enzymatic reactions (Nelson and Cox, 2008). Therefore, it is possible to note the effect of decreasing the oxidized forms of substrate in the presence of both an antioxidant agent, and PPO. Such an effect was observed in grape juice (Wu, 2014). The PPO effect on the oxidizing process could

be temporary eliminated in such a way. Antioxidant properties of glutathione (Kishkovsky and Scurihin, 1988) allow us to suggest that the anti-browning effect of this agent is due to its reduction action on the oxidized form of phenolic compounds.

The enzyme inhibitor must interfere with catalysis, and as a result, to slow or halt the enzymatic reaction. The data, gained by the scientists show that the same agent, such as glutathione, can be credited with having the properties of an inhibitor of oxidative enzyme activity in fruit products, as well as in meat (Gacche et al., 2004; Wu, 2014). These data call attention to the lack of specific activity of the agent, which is not well correlated with the classical theory of enzyme inhibition (Nelson and Cox, 2008). Biochemists use kinetic reaction studies to prove this. There are approaches for the control of PPO activity based on antisense techniques (Biord, 1991). X-ray crystallography and site-directed mutagenesis have been used to control PPO activity (Wagner and Benkovic, 1990). Classical methods based on analysing the Mixaelis-Menten equation can be used for investigating the activity of an agent as the enzyme inhibitor (Nelson and Cox, 2008). It was reported on high reproducibility of results of studying the enzymatic oxidative process with using the Mixaelis-Menten equation, and obtaining in such a way the data for offering the mechanism of reaction (Rogozhin and Peretolchin, 2010). The velocity of oxidized form conversion into reduced form with an antioxidant agent, credited with having PPO activity inhibition, should be checked, and compared with data from kinetic studies. The indirect methods in respect of using the antioxidant agents would not allow to obtain unambiguous results.

Matching the redox properties of fruit phenolic compounds with data according to which polyphenols can inhibit the free-radical processes of the oxidation of substrates

The possibility of phenolic compounds to be the inhibitors of free-radical oxidation of substrates was shown by many scientists (Azatyan *et al.*, 1973; Dinis *et al.*, 1994; Gerebin, 1994; Tang, 2001; Pazos, 2005; Lee *et al.*, 2006; Mielnik, 2006; Celiktar, 2007; Georgieva and Mihaylova, 2015).

The mechanism of the inhibition of free-radical oxidation of substrates in fruit beverages was represented (Gerebin, 1994) by the equation:

 RO_2 + PhO · \rightarrow products,

where RO_2 is a free radical, and PhO is a phenolic compounds.

 $\begin{cases} Q + H^+ \leftrightarrow HQ^+ - \text{chemical equilibrium;} \\ HQ^+ + \overline{e} \leftrightarrow HQ - \text{reaction of the transition} \\ 2HQ \leftrightarrow Q + H_2Q - \text{chemical equilibrium.} \\ \hline H_2Q \leftrightarrow Q + 2H^+ + 2\overline{e}, \end{cases}$

where Q is ortho-quinoid form of cyanidin; H_2Q is orthodiphenolic form of cyanidin; HQ is semiquinone.

Figure 2. The mechanism of redox conversion of anthocyans

The founding of semiquinone presence in quinone/hydroquinone system (Vetter, 1967), as well as our previous results, according to which there is a presence of semiquinone in the cyanidin reduced/ cyanidin oxidized system (Bocharova, 2008), allow us to offer the semiquinone form of diphenols as an inhibitor of free-radical oxidation of substrates in plant food products. Indeed, the mechanism of redox conversion of anthocyans was presented as shown in Figure 2 (Bocharova, 2008).

The mechanism of redox conversions of diphenols helps in understanding their positive role in the inhibition of free-radical processes of oxidation of substrates. Above mentioned do not deny the possibility of diphenols polymerisation. It is clear that reduced forms of diphenols cannot take part in such a process. As can be seen from the above, the presence of semiquinones in the equilibrium systems of plant flavonoids, such as anthocyans etc., and the chemical possibility, as well as the facts of inhibition the free-radical processes of oxidation of substrates with polyphenols, prove the vital role of semiquinones in this process.

Conclusion

(1) The two main theories, according to which the phenolic compounds in fruit products may be initiators of oxidation process, as well as its inhibitor, are the two sides of one redox phenomenon.

The phenolic compounds of redox-system of fruits with high reducing ability could act like antioxidants, and increase the stability of vitamin C; other phenolic compounds, with low reducing ability, like catechins, need to be reduced with such reducing agents as ascorbic acid or glutathione. The redoxpotential of polyphenols is the main factor, which effects the direction of non-enzymatic oxidative changes in fruit products. The value of standard redox-potential of polyphenolic substances can be the criterion for predicting the direction of redox conversions in fruit products.

To prove this, it is advisable to study the standard redox potentials of such polyphenols as

hesperidin, tangerine, and eriodictiol, by classical volt-amperometric method (Vetter, 1967). The redox-potentials of the above mentioned flavonoids should be less positive than the redox-potential of ascorbic acid/dehydroascorbic acid couple to prevent browning in fruit products, and than the potential of tartaric acid/dihydroxyfumaric acid couple, as well as of dihydroxyfumaric acid/ diketosuccinic acid couple to prevent the oxidation of tartaric acid.

(2) The functioning of flavonoids as redoxsystems are strictly connected with their ability to inhibit the free-radical processes of oxidation of substrates due to the semiquinone presence in the equilibrium system.

To prove this it is advisable to study the process of inhibition of the free-radical processes of oxidation of substrates with:

quinone/ hydroquinone system in which the presence of semiquinones can be neglected;

highly substituted quinones, such as tetramethyl-benzoquinone, in equilibrium systems of which there are substantial amounts of semiquinones (Vetter,1967).

(3) The glutathione can prevent browning of fruit products because of its low redox-potential, but not as inhibitor of PPO activity.

To prove this hypothesis, and investigate the activity of an agent with antioxidant properties as an enzyme inhibitor, the velocity of oxidized form conversion into reduced form, with such an agent, should be checked, and compared with data of kinetic studies for the resulting process. The classical methods based on analysing the Mixaelis-Menten equation are preferable.

References

- Adria, F. 2009. Modern Gastronomy: A to Z. Boca Raton: CRC Press.
- Akannia, O.O., Owumib, S.E. and Adaramoye, O.A. 2014. In vitro studies to assess the antioxidative, radical scavenging and arginase inhibitory potentials of extracts from *Artocarpus altilis, Ficus exasperate* and *Kigelia Africana*. Asian Pacific Journal of Tropical Biomedicine 4: S492–S499.
- Alper, N. and Acar, J. 2004. Removal of phenolic compounds in pomegranate juices using ultrafiltration and laccase-ultrafiltration combinations. Molecular Nutrition and Food Research 48(3): 184-187.
- Araji, S., Grammer, T.A., Gertzen, R., Anderson, S.D., Mikulic-Petkovsek, M., Veberic, R., Phu, M.L., Solar, A., Leslie, C.A., Dandekar, A.M. and Escobar, M.A. 2014. Novel Roles for the Polyphenol Oxidase Enzyme in Secondary Metabolism and the Regulation of Cell Death in Walnut. Plant Physiology 164(3): 1191–1203.

- Atawodi, S.E., Atawodi, J.C., Idakwo, P., Pfundstein, B., Haubner, R., Wurtele, G., Spiegelhalder, B., Bartsch, H. and Owen, R.W. 2009. Evaluation of polyphenolic composition and antioxidant activity of African variety of Dacryodes edulis. Journal of Medicinal Food 12(6): 1321-1325.
- Azatyan, N., Karpuhina, G., Belostockaya, I. and Komissarova, N. 1973. The mechanism of inhibition of oxidation processes with diphenols. Nephtehimiya 13(3): 435-440. (In Russian).
- Bahadoran, Z., Mirmiran, P. and Azizi, F. 2013. Dietary polyphenols as potential nutraceuticals in management of diabetes: a review. Journal of Diabetes and Metabolic Disorders 2(1): 43.
- Balasundrama, N., Sundramb, K. and Sammana, S. 2006. Phenolic compounds in plants and agri-industrial by-products: Antioxidant activity, occurrence, and potential uses. Food Chemistry 99(1): 191–203.
- Bharate, S.S. 2014. Non-enzymatic browning in citrus juice: chemical markers, their detection and ways to improve product quality. Journal of Food Science and Technology 51(10): 2271–2288.
- Bezusov, A., Telejenko, L. and Pilipenko, I. 2002. Decreasing the activity of plant phenols by methylation. Naukovi prazi ONAXT 23: 106-109. (In Russian).
- Bird, C.R. and Ray, J.A. 1991. Manipulation of Plant Gene Expression by Antisense RNA. Biotechnology and Genetic Engineering Reviews 9: 207-227.
- Block, G. and Patterson, B. 1992. Fruits, vegetables and cancer prevention: a review of the epidemiological evidence. Nutrition and Cancer 18: 1-29.
- Boekel, M., Fogliano, V., Pellegrini, N., Stanton, C., Scholz, G., Lalljie, S., Somoza, V., Knorr, D., Jasti, P.R. and Eisenbrand, G. 2010. A review on the beneficial aspects of food processing. Molecular Nutrition and Food Research 54(9): 1215–1247.
- Bocharova, O., Reshta, S. and Eshtokin, V. 2016. Toluene and benzyl alcohol formation in fruit juices containing benzoates. Journal of Food Processing and Preservation. [Accept], doi: 10.1111/jfpp.13054.
- Bocharova, O.V. 2008. Quality formation of anthocyancolored fruit juices. Odessa: Druk. (In Russian).
- Brownmiller, C., Howard, L.R. and Prior, R.L. 2008. Processing and Storage Effects on Monomeric Anthocyanins, Percent Polymeric Color, and Antioxidant Capacity of Processed Blueberry Products Journal of Food Science 73(5): 72-79.
- Caballero, B. 2013. Encyclopedia of Human Nutrition. 3rd ed. Oxford: Academic Press.
- Calado, J.C.P., Albertao, P.A., De Oliveira, E.A., Letra, M.H.S., Sawaya, A.C.H.F. and Marcucci, M.C. 2015. Flavonoid Contents and Antioxidant Activity in Fruit, Vegetables and Other Types of Food. Agricultural Sciences 6: 426-435.
- Celiktar, O.Y., Girgin, G., Orhan, H., Nichers, H.J., Bedir, E. and Sukan, F.V. 2007. Screening of free radical scavenging capacity and antioxidant activities of Rosmarinus officinalis extracts with focus on location and harvesting times. European Food Research and

Technology 24: 443-451.

- Cespedes, C.L. 2008. Antioxidant and cardioprotective activities of phenolic extracts from fruits of Chilean blackberry *Aristotelia chilensis* (Elaeocarpaceae), Maqui. Food Chemistry 107(2): 820-829.
- Gharras, H.E. 2009. Polyphenols: food sources, properties and applications – a review. International Journal of Food Science and Technology 44(12): 2512–2518.
- Cheynier, V. 2012. Phenolic compounds: from plants to foods. Phytochemistry Reviews 11(2): 153-177.
- Choe, E. and Min, D.B. 2009. Mechanisms of Antioxidants in the Oxidation of Foods. Comprehensive Reviews in Food Science and Food Safety 8(4): 345–358.
- Dai, J. and Mumper, R.J. 2010. Plant Phenolics: Extraction, Analysis and Their Antioxidant and Anticancer Properties. Molecules 15: 7313-7352.
- Dinis, T.C., Madeira, V.M. and Almeida, L.M. 1994. Action of phenolic derivatives (acetaminophen, salicylate, and 5-aminosalicylate) as inhibitors of membrane lipid-peroxidation and as peroxyl radical scavengers. Archives of Biochemistry and Biophysics 315: 161-169.
- Du, G., Li, M. and Ma, F. 2009. Antioxidant capacity and the relationship with polyphenol and Vitamin C in Actinidia fruits. Food Chemistry 113(2): 557–562.
- Dyidogan, N. and Bayindirli, A. 2004. Effect of L-cysteine, kojicacid and 4-hexylresorcinol combination on inhibition of enzymatic browning in Amasya apple juice. Journal of Food Engineering 62: 299-304.
- El Hosry, L., Auezova, L., Sakr, A. and Hajj Moussa, E. 2009. Browning susceptibility of white wine and antioxidant effect of glutathione. International Journal of Food Science and Technology 44: 2459-2464.
- Fedina, P.A., Yashin, A.Ja. and Chernousova, N.I. 2010. Determination of antioxidants in products of plant origin by amperometric method. Himija rastitelnogo syrija 26(2): 91–97. (In Russian).
- Flora, S.J.S. 2009. Structural, chemical and biological aspects of antioxidants for strategies against metal and metalloid exposure. Oxidative Medicine and Cellular Longevity 2(4): 191–206.
- Flaumenboum, B.L, Bezusov, A.T., Storojuk, V.N. and Xomich, G.P. 2006. The physicochemical base of canned food production. Odessa: Astroprint. (In Ukrainian).
- Gacche, R.N., Warangkar, S.C. and Ghole, V.S. 2004. Glutathione and cinnamon acid: Natural dietary components used in preventing the process of browning by inhibition of polyphenol oxidase in apple juice. Journal of Enzyme Inhibition and Medicinal Chemistry 19: 175-179.
- Georgieva, L. and Mihaylova, D. 2015. Screening of total phenolic content and radical scavenging capacity of Bulgarian plant species. International Food Research Journal 22(1): 240-245.
- Gerebin, Y. 1994. The mechanism of non-enzymatic oxidation in fruit beverages. Naukovi pratzi ODAXT 15: 47-52. (In Russian).
- Girones-Vilaplana, A. 2012. A novel beverage rich in antioxidant phenolics: Maqui berry (Aristotelia

chilensis) and lemon juice. LWT-Food Science and Technology 47(2): 279-286.

- Goronovsky, I.T., Nazarenko, Y.P. and Nekryach, E.F. 1987. Short Guide to Chemistry. Kiev: Naukova Dumka. (In Russian).
- Handique, J.B. and Baruah, J.G. 2002. Polyphenolic compounds: an overview. Reactive and Functional Polymers 52(3): 163–188.
- Heinonen, M.I. 2007. Antioxidant activity and antimicrobial effect of berry phenolics – a Finnish perspective. Molecular Nutrition and Food Research 51(6): 684-691.
- Housecroft, C.E. and Constable, E.C. 2006. Chemistry. Harlow: Pearson Education Limited.
- Hui, H.Y. 2006. Handbook of fruit and fruit processing. Ames: Blackwell Publishing.
- Hui, H.Y. and Evranuz, E.O. 2016. Handbook of vegetable preservation and processing. Boca Raton: CRC Press.
- Huang, W.Y., Cai, Y.Z. and Zhang, Y. 2010. Natural phenolic compounds from medicinal herbs and dietary plants: potential use for cancer prevention. Nutrition and Cancer 62(1):1-20.
- Hunt, M.D., Eannetta, N.T., Yu, H., Newman, S.M. and Steffens, J.C. 1993. cDNA cloning and expression of potato polyphenol oxidase. Plant Molecular Biology 21(1): 59-68.
- Hutchings, J.B. 1994. Food Colour and Appearance. Glasgow: Springer Science + Business Media Dordrecht.
- Ignat, I., Volf, I. and Popa, V.I. 2011. A critical review of methods for characterisation of polyphenolic compounds in fruits and vegetables. Food Chemistry 126(4): 1821–1835.
- Ivanova, D., Gerova, D., Chervenkov, T. and Yankova, T. 2005. Polyphenols and antioxidant capacity of Bulgarian medicinal plants. Journal of Ethnopharmacology 69: 145-150.
- Jang, J.H. and Moon, K.D. 2011. Inhibition of polyphenol oxidase and peroxidase activities on fresh-cut apple. Food Chemistry 124: 444–449.
- Kasikci, M.B. and Bagdatlioglu, N. 2016. High hydrostatic pressure treatment of fruit, fruit products and fruit juices: a review on phenolic compounds. Journal of Food and Health Science 2(1): 27-39.
- Katalinic, V., Milos, M., Kulisic, T. and Jukic, M. 2006. Screening of 70 medicinal plants for antioxidant capacity and total phenols. Food Chemistry 94: 550-557.
- Keerthi, M., Prasanna, J.L., Aruna, M. S. and Rao, N. R. 2014. Review on polyphenols as natures gift. World Journal of Pharmacy and Pharmaceutical Sciences 3(4): 445-455.
- Kishkovsky, Z.N. and Skurihin, I.M. 1988. Chemistry of wine. Moscow: Agropromizdat. (In Russian).
- Kolodziejczyk, K., Milala, J., Sojka, M., Kosmala, M. and Markowski, J. 2010. Polyphenol oxidase activity in selected apple cultivars. Journal of Fruit and Ornamental Plant Research 18(2): 51-61.
- Korotkova, E.I., Voronova, O.A. and Dorozhkov, E.V. 2012. Study of antioxidant properties of flavonoids by

voltammetry. Journal of Solid State Electrochemistry 16(7): 2435-2440.

- Kratchanova, M., Denev, P., Ciz, M., Lojek, A. and Mihailov, A. 2010. Evaluation of antioxidant activity of medicinal plants containing polyphenol compounds. Comparison of two extraction systems. Acta Biochimica Polonica 57 (2): 229-234.
- Krylov, S.N., Krylova, S.M. and Rubin, L.B. 1993. Mechanism of inhibition of enzymatic oxidation of indole-3-acetic acid by phenols. Biochimiya 58(6): 953-961.
- Lee, C.H., Reed, J.D. and Richards, M.P. 2006. Ability of various polyphenolic classes from cranberry to inhibit lipid oxidation in mechanically separated turkey and cooked ground pork. Journal of Muscle Foods 17(3): 248-266.
- Leopoldini, M., Russo, M. and Toscano, M. 2011. The molecular basis of working mechanism of natural polyphenolic antioxidants. Food Chemistry 125(2): 288-306.
- Manach, C., Scalbert, A., Morand, C., Remsy, C. and Jimenez, L. 2004. Polyphenols: food sources and bioavailability. American Journal of Clinical Nutrition 79(5): 727-747.
- Marh, Z., Dmitrieva, E. and Evstigneev, K. 1985. Canned food for babies and children. Moscow: Agropromizdat. (In Russian).
- Martiner, M.V. and Whitaker, J.R. 1995. The biochemistry and control of enzymatic browning. Food Science and Technology 6(60):195-200.
- Mertz, C., Cheynier, V., Günata, Z. and Brat, P. 2007. Analysis of phenolic compounds in two blackberry species (*Rubus glaucus* and *Rubus adenotrichus*) by high-performance liquid chromatography with diode array detection and electrospray ion trap mass spectrometry. Journal of Agriculture and Food Chemistry 55: 8616-8624.
- Mielnik, M.B.2006. Grape seed extract as antioxidant in cooked, cold stored turkey meat. LWT-Food Science and Technology 39(3): 191-198.
- Murillo, E., Britton, G.B. and Durant, A.A. 2012. Antioxidant activity and polyphenol content in cultivated and wild edible fruits grown in Panama. Journal of Pharmacy and Bioallied Sciences 4(4): 313–317.
- Nelson, D.L. and Cox, M.M. 2008. Principles of biochemistry. New York: W.H. Freeman and company.
- Nikolaevskiy, A.N., Kycher, R.V. and Filipenko, T.A. 1978. Kinetic and the mechanism of liquid-faze inhibition of hydrocarbons with diphenols, p. 103-119. Kiev: Naykova dumka, (In Russian).
- Osuga, D.A. Van Der Schaaf, Whitaker, J. R. 1994. Control of polyphenol oxidase activity using a catalytic mechanism. In Yada, RY., Jackman, R.L.and Smith, S.L. (Eds). Protein Structure-Function Relationships in Foods, p. 62-88. New York: Springer US.
- Pastrana-Bonilla, E., Akoh, C.C, Sellappan, S. and Krewer, G. 2003. Phenolic content and antioxidant capacity of Muscadine grapes. Journal of Agriculture and Food Chemistry 51: 5497-5503.

- Pazos, M. 2005. Activity of grape polyphenols as inhibitors of the oxidation of fish lipids and frozen fish muscle. Food Chemistry 92(3): 547-557.
- Pizzocaro, F., Torregiani, D. and Gilardi, G. 1993. Inhibition of apple polyphenoloxidase (PPO) by ascorbic acid, citric acid and sodium chloride. Food Process Preservation 17: 21–30.
- Preedy, V. 2014. Processing and Impact on Antioxidants in Beverages. Oxford: Academic Press.
- Ramful, D., Tarnus, E., Aruoma, O.I., Bourdon, E. and Bahorun, T. 2011. Polyphenol composition, vitamin C content and antioxidant capacity of Mauritian citrus fruit pulps. Food Research International 44(7): 2088– 2099.
- Ramaswamy, H. and Marcotte, M. 2005. Food Processing: Principles and Applications. London: CRC Press.
- Ren, W., Qiao, Z., Wang, A. and Zhang, L. 2003. Flavonoids: Promising anticancer agents. Medical Research Reviews 23: 519-534.
- Rogozhin, V.V. and Peretolchin, D.V. 2010. Kinetics of oxidation of ascorbic acid by horseradish peroxidase. Vestnik Yujno-Yralskogo Universiteta 11: 61-65.
- Sasikumar, J.M, Poulin, R.C., Meseret, C.E. and Selvakumar, P. 2015. In vitro analysis of antioxidant capacity of Indian yellow raspberry (*Rubus ellipticus* Smith). International Food Research Journal 22(4): 1338-1346.
- Scoricova, Y.G. 1973. Polyphenols of fruit and berries, and colour formation of products. Moscow: Pischevaya promuslennost. (In Russian).
- Sharma, K.D., Kaushal, M. and Kaushal, B.B.L. 2002. Canning of Peach-halves in Fruit Juice. Journal of Scientific and Industrial Research 61: 823-827.
- Siddiq, M. and Cash, J.N. 2000. Physico-chemical properties of polyphenol oxidase from d'Anjou and Bartlett pears (*Pyrus communis* L.), Journal of Food Processing 24(5): 353-364.
- Sioud, F.T. and Luh, B.S. 1966. Polyphenolic compounds in pear puree. Food Technology 20(4): 182-186.
- Siriwoharn, T., Wrolstad, R.E., Finn, C.E. and Pereira, C.B. 2004. Influence of cultivar, maturity, and sampling on blackberry (*Rubus* L. Hybrids) anthocyanins, polyphenolics, and antioxidant properties. Journal of Agriculture and Food Chemistry 52: 8021-8030.
- Sripakdee, T., Sriwicha, A., Jansam, N., Mahachai, R. and Chanthai, S. 2015. Determination of total phenolics and ascorbic acid related to an antioxidant activity and thermal stability of the Mao fruit juice. International Food Research Journal 22(2): 618-624.
- Suh, H.J. and Park, S. 2011. Inhibition of browning of fresh apple juices by natural phytochemicals from *Rumex crispus* L. Seed. Journal of the Korean Society for Applied Biological Chemistry 54: 524-530.
- Tang, S. 2001. Anti-oxidant activity of added tea catechins on lipid oxidation of raw minced red meat, poultry and fish muscle. International Journal of Food Science and Technology 36(6): 685-692.
- Thutnham, D. 1992. Functionally important antioxidants and free radical scavengers in foods. Food Science and Technology Today 6(1): 42-46.

- Tressler, D.K. and Jocelyn, M.A. 1957. Chemistry and Technology of Berry Fruit and Vegetable Juices. Moscow: Pishchepromizdat.
- Tsao, R. 2010. Chemistry and Biochemistry of Dietary Polyphenols. Nutrients 2: 1231-1246.
- Vamos, V.L. 1981. Polyphenol oxidase and peroxidase in fruits and vegetables. Food Science 15: 49–127.
- Van Acker, S.A., Den Berg, D.J., Tromp, M.N., Griffioen, D.H., Van Bennekom, W.P. and Van der Vijgh, W.J. 1996. Structural aspects of an antioxidant activity of flavonoids. Free Radical Biology and Medicine 20: 331-342.
- Vetter, J.K. 1967. Electrochemical kinetic. Moscow: Ximiya.
- Vitti, M.C.D., SasakiI, F.F., MiguelI, P., KlugeI, R.A. and Moretti, C.L. 2011. Activity of enzymes associated with the enzymatic browning of minimally processed potatoes. Brazilian Archives of Biology and Technology 54(5): 983-990.
- Watson, R.R, Preedy, V.R. and Zibadi, S. 2014. Polyphenols in Human Health and Disease. San Diego: Academic Press.
- Wagner, C.R. and Benkovic, S.J. 1990. Site directed mutagenesis: a tool for enzyme mechanism dissection. Trends Biotechnology 8: 263-270.
- Wojdylo, A., Oszmiansky, J. and Czemerys, R. 2007. An antioxidant activity and phenolic compounds in 32 selected herbs. Food Chemistry 105: 940-949.
- Wu, S.J. 2013. Inhibition of enzymatic browning of the meat of *Clanis bilineata* (Lepidoptera) by glutathione. Food Science and Technology Research 19: 347-352.
- Wu, S.J. 2014. Glutathione suppresses the enzymatic and non-enzymatic browning in grape juice. Food Chemistry 160: 8-10.
- Yoruk, R. and Marshall, M.R. 2003. Physicochemical properties and function of plant polyphenol oxidase: A review. Journal of Food Biochemistry 27: 361-422.
- Zaprometov, M. 1964. Biochemistry of catechins. Moscow: Nauka. (In Russian).
- Zheng, W. and Wang, Y. 2001. Antioxidant activity and phenolic com-pounds in selected herbs. Food Chemistry 49: 5165-5170.